

TRANSFECTION OF HUMAN NEUROBLASTOMA SH-SY5Y CELLS AND MONKEY COS-7 CELLS WITH METAFECTENE PRO

Vanessa de la Rosa, Francisco J. Alonso, Carolina Lobo and Javier Márquez.

Department of Molecular Biology and Biochemistry, Faculty of Sciences, University of Malaga, 29071 Malaga (SPAIN)

Materials

Metafectene PRO, a polycationic liposomal transfection reagent, was obtained from Biontex Laboratories GmbH (Munich, Germany). The cell growth culture media (RPMI-1640 and Minimum Essential Medium Eagle), penicillin and streptomycin solutions were obtained from Sigma.

The plasmid, pEGFP-C1 (PT3027-5, Clontech Laboratories), encoding green fluorescent protein was used for evaluating transfection efficiency.

Cells

The human neuroblastoma cells SHSY-5Y were cultured in Minimum Essential Medium Eagle supplemented with 10% Fetal Bovine Serum (Sigma), anphotericin (1.25 mg/mL) penicillin (100U/mL) and streptomycin (100U/mL). The COS-7 cells were cultured in RPMI medium with the same supplements. The growth conditions were 37°C and 5% of CO₂.

Transfection protocol

For transfection, SHSY-5Y (1.5 x 10⁵ cells/well) and COS 7 cells (2 x 10⁵ cells/well) were seeded in 1 mL of serum free medium (OPTI-MEM[®], Gibco) in a 12-well microtitter plate, and then incubated at 37°C in a 5% CO₂ incubator overnight to obtain 80-90% confluence. Cells were pre-washed with serum-free OPTI-MEM[®] medium and covered with 1 mL of the same medium. Metafectene PRO was complexed with the pEGFP-C1 plasmid at reagent:DNA (μL:μg) ratios of 1:0.5, 2:0.5, 4:0.5, 6:0.5, 2:1, 4:1, 8:1, 12:1, 4:1.5, 8:1.5, 12:1.5 or 16:1.5. Complexes were prepared by mixing Metafectene PRO with 0.1 mL of serum-free OPTI-MEM[®] medium, followed by the addition of plasmid DNA. The mixture was incubated for 15 min at room temperature after addition of transfection reagent, and another 15 min after addition of DNA. Metafectene PRO complexes with DNA were added in a volume of 0.1 mL per well and cells were incubated for 24 h at 37°C in a 5% CO₂ incubator. Then cells were washed three times with sterile PBS, added 1 mL of fresh PBS and analyzed under UV light in a Nikon microscope (Eclipse E 800).

Results

Metafectene PRO was complexed with the pEGFP-C1 plasmid at reagent:DNA (μ L: μ g) ratios of 1:0.5, 2:0.5, 4:0.5, 6:0.5, 2:1, 4:1, 8:1, 12:1, 4:1.5, 8:1.5, 12:1.5 or 16:1.5. In case of COS-7 cells, all ratios showed transfection, but the best ratios were 2:0.5, 4:0.5, 8:1, 12:1 and 12:1.5, obtaining 80-95% of transfection level. Neuroblastoma cells were no transfected with ratios 4:0.5, 8:1 and 4:1.5. The transfection percentages are shown in Figure 1. Figure 2 and 3 show transfected COS7 and SHSY-5Y cells.

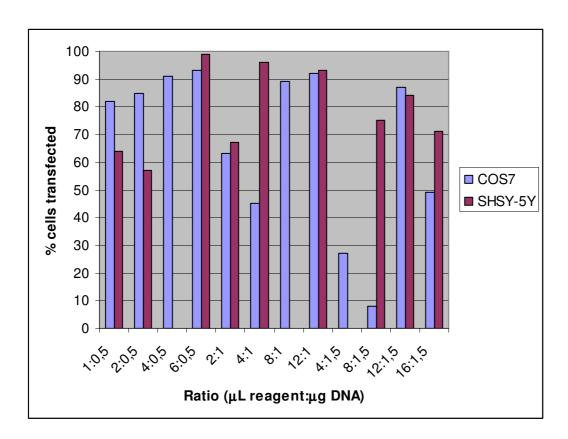


Figure 1. Percentage of cells transfected with pEGFP-C1 using Metafecten-PRO. Two differents cell lines, COS7 and neuroblastoma cells (SHSY-5Y) were transfected with different ratios of Metafecten-PRO/DNA. Approximately in 70% of the experiments we observed a transfection level higher than 50%.

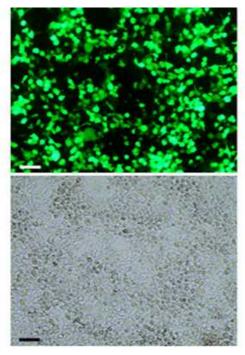


Figure 2. The above image shows UV microscopy of COS7 cells transfected with pEGP-C1 using Metafectene-PRO. The ratio used was 4 ml reagent/0.5 μg DNA. Image shows optical microscopy of the same cells. Scale bars are equivalent to $100 \, \mu m$.

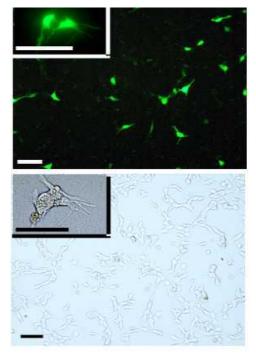


Figure 3. Neuroblastoma cells transfected with pEGP-C1 using Metafecten-PRO. The ratio was 2 ml reagent/0.5 μg DNA. Top: UV microscopy of neuroblastoma cells. Bottom: optical microscopy of the same cells. The boxes show some cells detailed. Scale bars are equivalent to 100 μm.